

Measurement of Cell-Free RNA Signatures After Severe Trauma in a Porcine Model Supports Future Platelet Transcriptomic Profiling

Teryn R. Roberts^{1*}, Alexander Fields^{2*}, Roland J. Bainton², Fahima Mayer², Yale Santos², Cedric Bainton², Karl Zoghbi², Nasima Mayer², Kaitelynn B. Beely¹, Brendan M. Beely¹, George T. Harea¹, Daniel S. Wendorff¹, Lawrence A. Renna¹, Andriy I. Batchinsky¹, Lucy Z. Kornblith²

¹Autonomous Reanimation and Evacuation Research Institute, The Geneva Foundation, San Antonio, TX, USA; ²University of California, San Francisco, San Francisco, CA, USA

The opinions or assertions contained herein are the private views of the author and are not to be construed as official or as reflecting the views of U.S. Government, the Department of Defense, or The Geneva Foundation.

Introduction

- Thromboinflammation – a complex dysregulation of coagulation, inflammation, and immune function – drives downstream complications following combat and major civilian trauma.
- Platelets play a central role in thrombosis and inflammation. Emerging evidence from platelet transcriptomics demonstrates that platelets harbor dynamic, disease-specific RNA signatures that reflect and actively modulate thrombinflammation.
- Platelet-based functional assays have shown inconsistent associations with thrombinflammatory outcomes after trauma, limiting utility in biomarker development and therapeutic discovery.
- Evaluation of platelet RNA signatures in clinical practice is challenged by logistics - including timing of sample collection, lack of pre-injury assessment, and heterogeneity of injury mechanism intervention. Multi-day translational research studies offer a unique opportunity for protocolized investigation.
- ~30% of plasma cell-free RNA (cfRNA) is platelet-derived, offering a unique opportunity to interrogate platelet-related molecular responses.

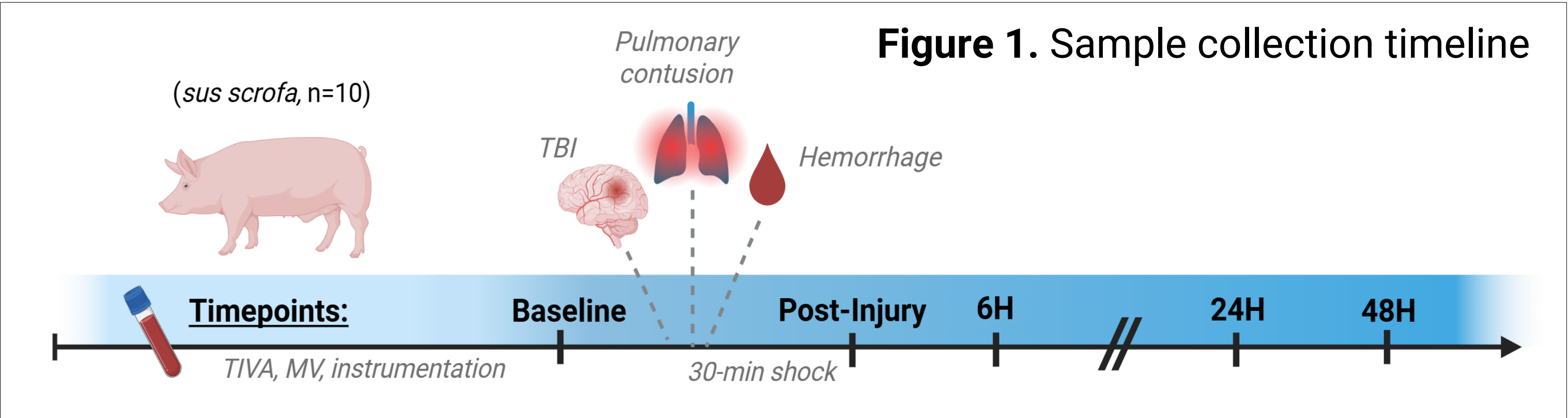
Objective and Hypothesis

Objective: Interrogate plasma space for platelet-related cfRNA in serial samples in a combat-relevant swine polytrauma model with 48-hour follow-up.

Hypothesis: cfRNA profiles demonstrate consistent time-dependent changes following trauma.

Methods

- Yorkshire swine (*sus scrofa domesticus*; n=10, 54±2 kg) were anesthetized, mechanically ventilated, and instrumented.
- Following baseline data collection, swine encountered traumatic brain injury, bilateral pulmonary contusion, and controlled hemorrhage; followed by 30-min shock period.

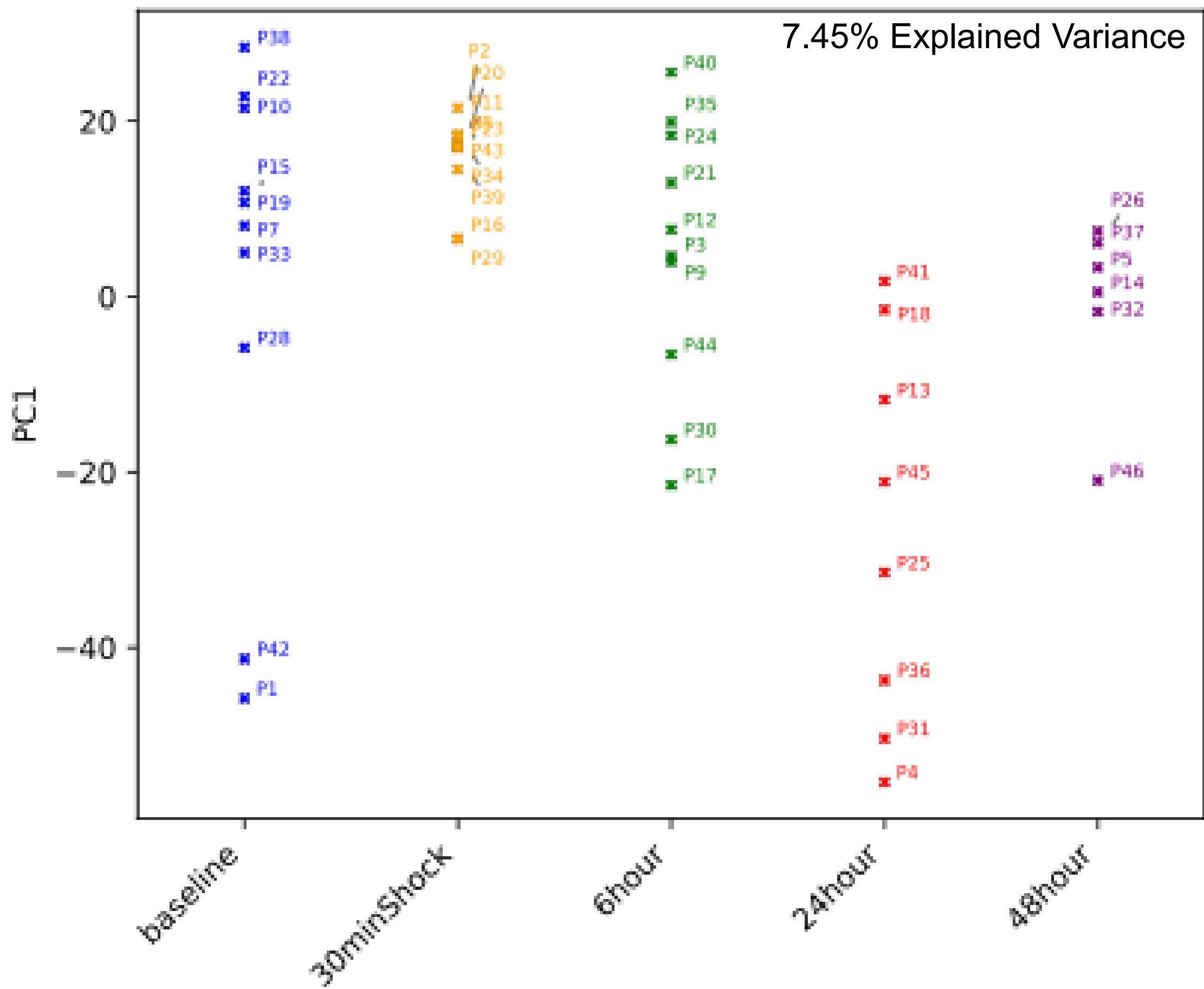


Methods

- Citrated plasma was prepared at discrete timepoints (**Figure 1**) and flash frozen (-80 °C).
- cfRNA was extracted and subjected to next-generation sequencing (n=43 passed QC; ~1M reads / sample). Fibrinogen was quantified (Stago CompactMax).
- EdgeR was used for differential gene discovery. Principal component analysis (PCA) was performed on normalized counts per million to examine patterns of gene discovery between individuals and timepoints.
- ANOVA or paired T-tests were used to assess differences in principal components at timepoints (p<0.05 for significance).
- Pearson’s correlation was performed on principal components and fibrinogen level.

Results

- cfRNA profiling revealed consistent platelet-related genomic response following polytrauma, despite baseline inter-animal variability (**Figure 2**).
- Initial post-injury timepoint showed strongest agreement between samples, suggesting initial trauma perturbation drives samples to similar projection status (**Figure 2**).



	p-value vs Baseline	p-value vs Post-Injury	p-value vs 6 H	p-value vs 24 H
Post-Injury	0.14	---	---	---
6H	0.67	0.03	---	---
24H	0.019	.0017	0.016	---
48H	0.56	0.04	0.27	0.015

Figure 2. PCA showed trauma-related genomic patterns (PC1). Paired T-tests highlight significantly different timepoints.

Results

- One genomic pattern (PC10) was identified with significant changes between timepoints (ANOVA p=0.003) suggesting a stereotyped early response to injury, with modulation over 48 hours. This pattern was correlated with fibrinogen levels (**Figure 3**).
- Cumulative findings suggest that trauma induces a structured, reproducible cfRNA signature in circulation, and is correlated with other physiological measures.

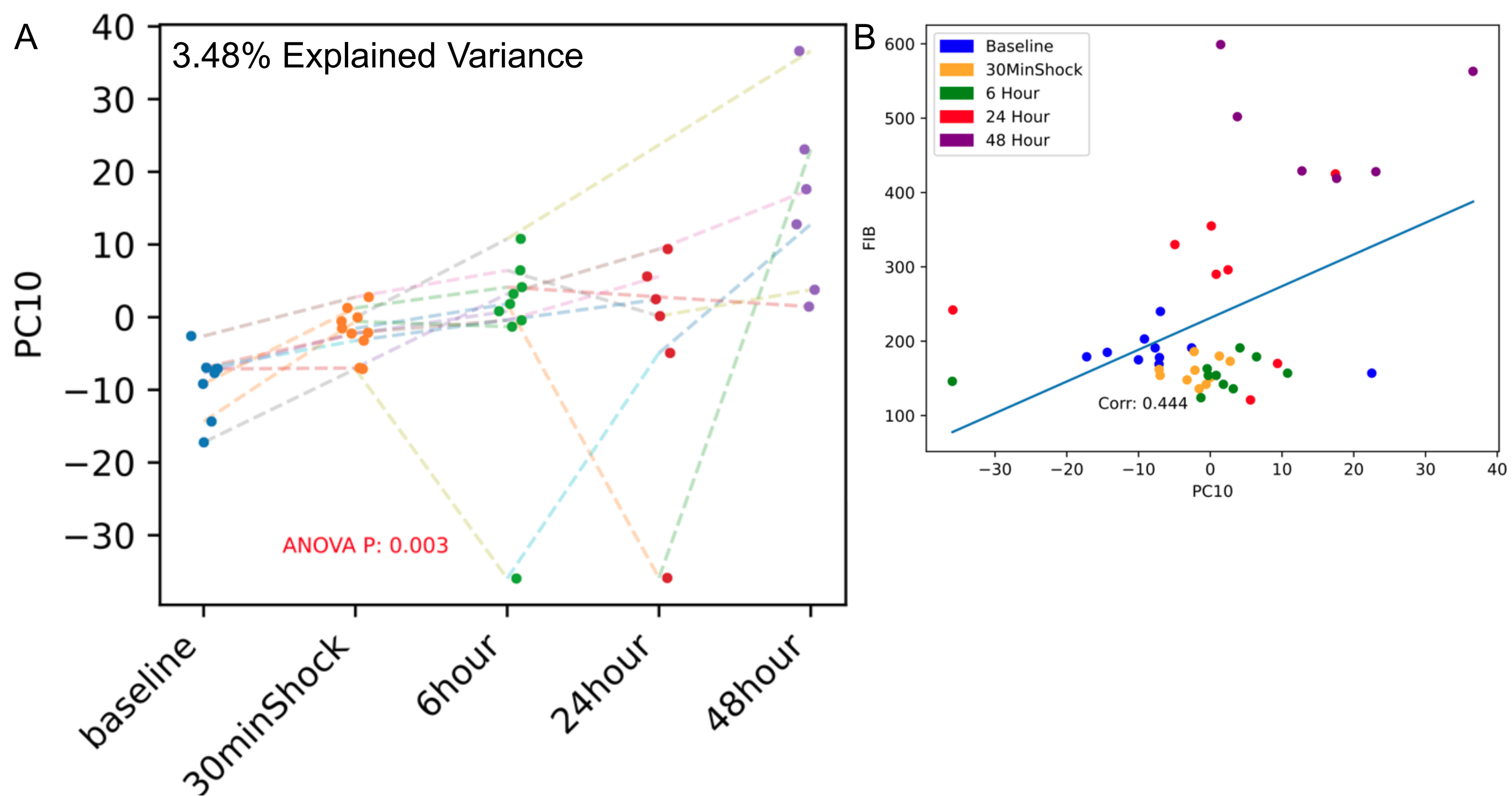


Figure 3. (A) Principal component 10 differed significantly (p=0.003) over time. (B) PC10 was correlated with fibrinogen level (Pearson’s correlation coefficient 0.444).

Conclusions

- Pilot data demonstrate that platelet-related cfRNA captures coherent, time-dependent molecular responses to severe trauma in a translational model.
- cfRNA PCA analysis may discover different cell types undergoing lysis, restoration, and adaptation over the injury course – necessitating future analysis of cell-resolved analysis of gene expression changes in acute trauma.
- These findings support isolating platelets for dedicated platelet transcriptomics, enabling novel biomarker identification and therapeutic target discovery for military trauma care.

Acknowledgments

Animal studies were supported via the Defense Medical Research and Development Program (Award No. W18XWH-19-2-0008 (PI Andriy Batchinsky, MD) and administered through The Geneva Foundation, Tacoma, WA. Genomic analyses were supported via the National Institute of Health National Institute of General Medical Sciences (R35GM150656, PI Lucy Kornblith, MD) and the American College of Surgeons (ACS; PI, Lucy Kornblith). Opinions, interpretations, conclusions and recommendations are those of the authors and are not necessarily endorsed by the DOW, NIH, or ACS. In conducting research using animals, investigators adhered to laws of the United States and regulations of the Department of Agriculture.