

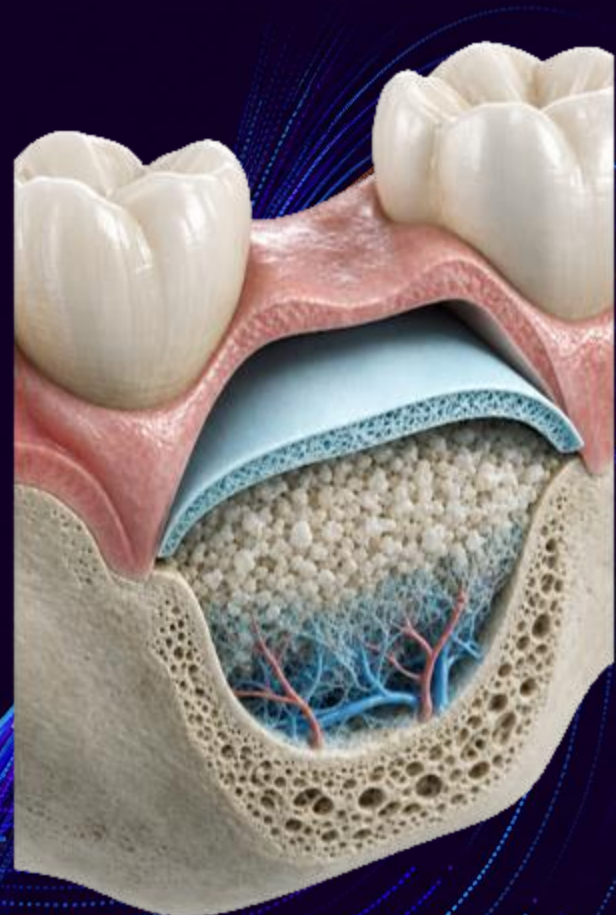
Translational Evaluation of Novel Amino-acid based Membrane for GBR

Designable microarchitecture, preclinical
performance, and operational relevance

MHSRS-26-4553 · CRITICAL CONCERNS IN OPERATIONAL DENTISTRY

Sherif Soliman, PhD, MBA

Matregenix, Inc. · MHSRS 2026



Disclosure, Investigational Status, and Disclaimer



Financial disclosure

Dr. Sherif Soliman is Founder and Chief Executive Officer of Matrogenix, Inc. and has an equity and financial interest in the PEU membrane technology discussed.



Investigational technology

The barrier membrane is investigational and is not cleared by the U.S. Food and Drug Administration. This presentation includes preclinical, in vitro, and company-supplied feasibility data.



Required disclaimer

The views expressed are those of the presenter and do not necessarily reflect the official policy of the Department of Defense, Defense Health Agency, Uniformed Services University, National Institutes of Health, or the U.S. Government. Mention of commercial products does not imply endorsement.

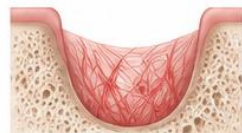
Barrier Membranes: The Foundation of Guided Bone Regeneration



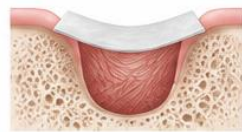
Bone defect



Soft tissue invasion



Bone regeneration fails



Barrier membrane placed



Guided Bone Regeneration

- ✓ Space maintenance
- ✓ Bone cells repopulate
- ✓ New bone formation

Five classic design criteria (Scantlebury, 1993)

Biocompatibility

Low tissue reactivity; avoids inflammation-driven dehiscence

Cell occlusivity

Blocks soft-tissue cell ingrowth into the compartment

Space maintenance

Resists collapse to preserve regenerative volume

Tissue integration

Anchors to host tissue and supports vascular contact

Clinical manageability

Handles, conforms, and stays stable when wet

Optimizing one criterion usually forces a compromise in another — no single existing material wins on all five.

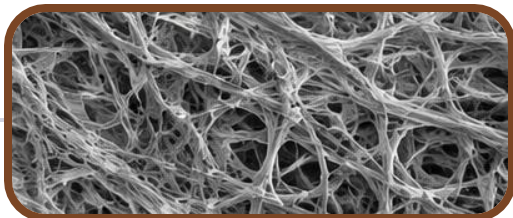
Why Do We Still Need Better GBR Membranes?

Current Barrier Membranes



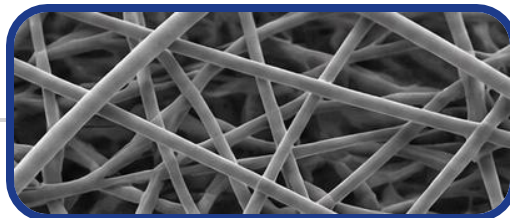
COLLAGEN MEMBRANES

- ✓ Excellent biocompatibility
- ✓ Native ECM-like structure
- ✗ Low mechanical strength
- ✗ Rapid degradation
- ✗ Animal-derived variability



SYNTHETIC MEMBRANES

- ✓ Better mechanical properties
- ✓ Controlled manufacturing
- ✗ Acidic degradation products
- ✗ Potential inflammatory response
- ✗ Difficult to handle



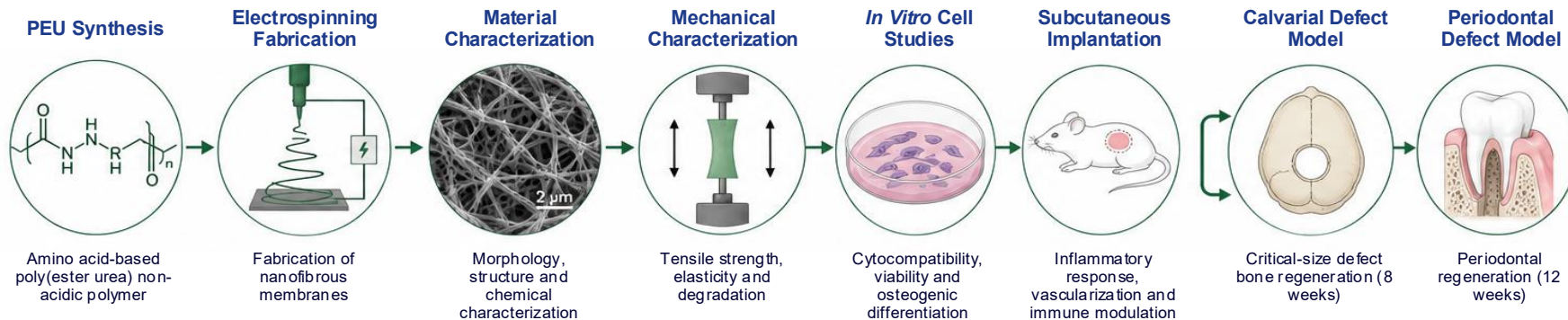
Study Hypothesis and Design



Study Hypothesis

“An electrospun amino acid–based PEU barrier membrane will achieve non-inferior biocompatibility and bone-regenerative performance compared with clinically used collagen membranes, while demonstrating superior handling characteristics and a more favorable cost–performance profile”

Study Design



Electrospinning: A Scalable, Animal-free Manufacturing Route



Minutes, not days

A 1 m² synthetic sheet is produced in minutes; equivalent donor tissue takes days to weeks to source and process.



Why it matters operationally



Scalable

Roll-to-roll deposition supports reproducible, higher-throughput production.



Cost-effective

Synthetic feedstock avoids costly tissue sourcing, purification, and screening.



Animal-free

No xenogeneic or donor material — no associated ethical or disease-transmission concerns.

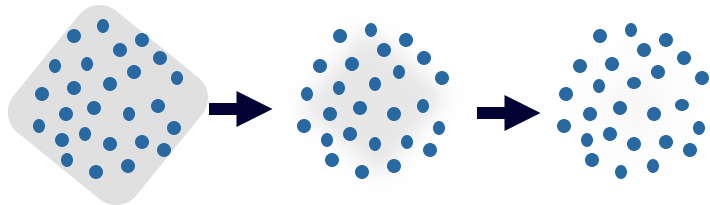
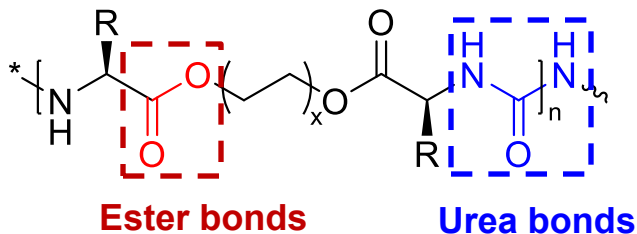


Tunable

Fiber-scale control of mechanics, porosity, and degradation for a target indication.

Amino-acid-based poly(ester urea): A Novel Barrier Chemistry

Amino Acid-Based Poly(ester urea)s



Uniform degradation & Non-acidic byproducts

Progress in Polymer Science **2024**, 156, 101866.

Advantages

Versatile thermoplastic

- Many validated form factors
- Film, tube, suture, mesh, scaffold

Synthetic Flexibility

- Tunable mechanical properties
- Customizable degradation (weeks to months)

Biocompatible

- Non-toxic byproducts
- Limited inflammatory response

Controlled drug delivery

- Drug stabilization
- Uniform and sustained (days to weeks)

Technology and Material Platforms Created MatriNova

MatriNova: Electrospun PEU Platform for Tissue Repair

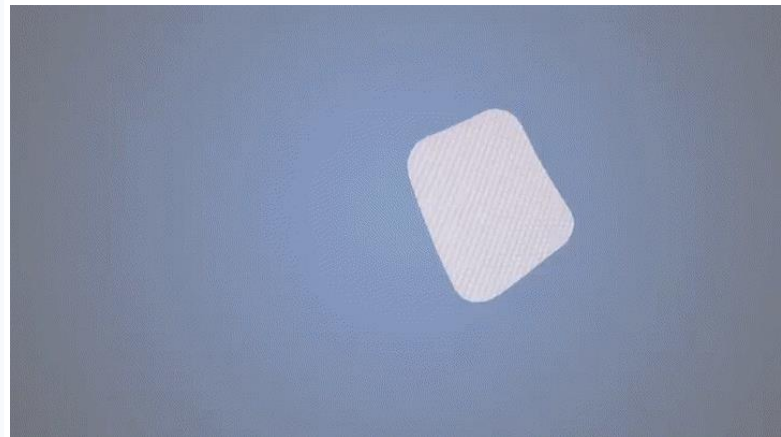
MATRINOVA

SYNTHETIC MEMBRANE

A synthetic bilayer barrier membrane designed for GTR and GBR applications

Animal-free — no disease-transmission risk
Easier to handle
Designed for lower cost than biologics

SAFER • Easier Handling • LOWER COST



One Material, Multiple Operational Indications

The amino-acid PEU platform is programmed at the fiber scale — thickness, pore size, and degradation tuned per indication



Dental GBR

Excludes soft tissue; guides alveolar bone and cementum regeneration.

Pre-clinical complete



Wound healing

Full-thickness wound matrix; ECM-mimetic, antimicrobial support.

Pre-clinical complete



Rotator cuff

Reinforcement scaffold for load-bearing tendon repair.

ovine feasibility



Ophthalmic

Synthetic "amniotic membrane" for the ocular surface.

program-stage



Dural repair

Watertight barrier for neurosurgical defects.

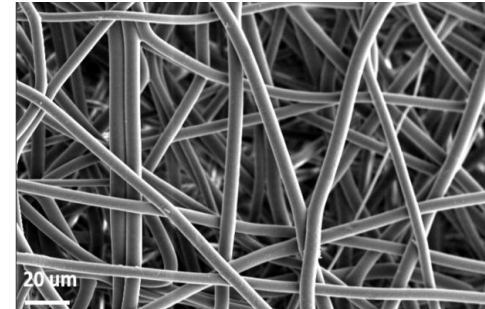
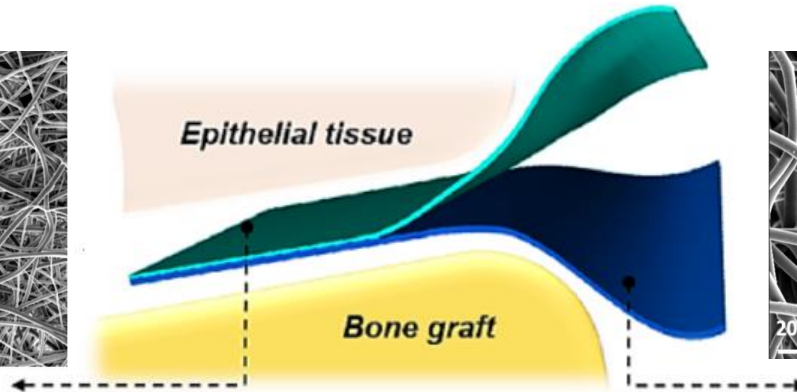
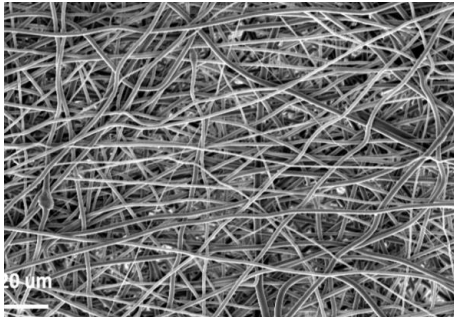
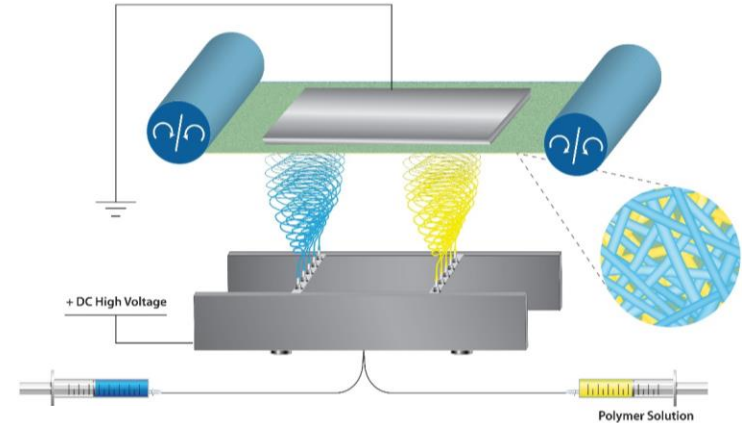
concept-stage

ONE CHEMISTRY · TUNED ARCHITECTURE · TISSUE FUNCTION

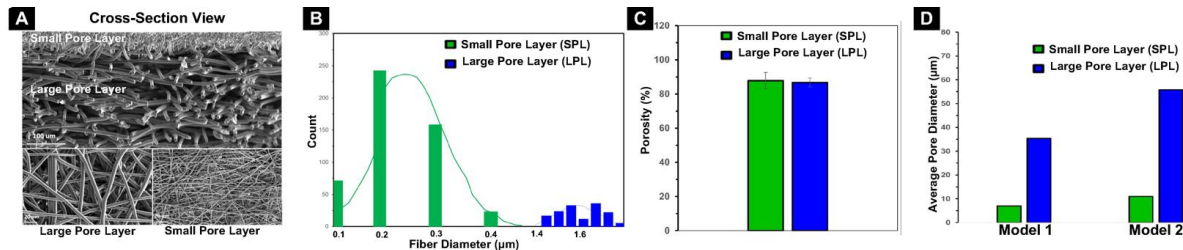
Electrospun PEU Bilayer Membrane Design

Bilayer Design

- **Small pore layer (SPL):** achieve the prevention of epithelial tissue ingrowth
- **Large pore layer (LPL):** encourage cell infiltration and guide bone tissue formation



The Bilayer Architecture Separates Tissue Exclusion from Osteogenic Support

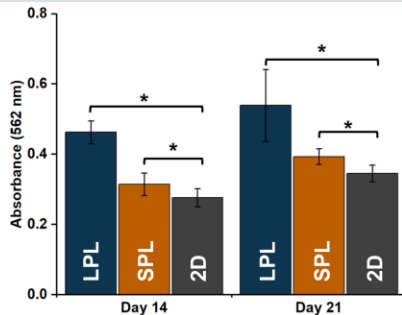


>90% porosity

in both small- and large-pore layers

≤10 / ≥50 μm

target pore regimes for barrier and regenerative faces



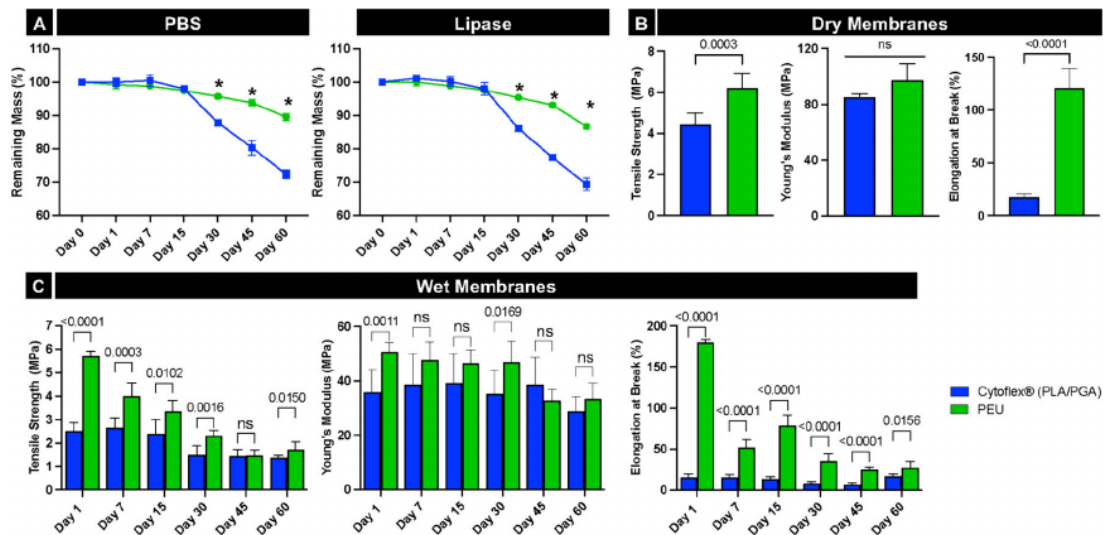
The large-pore layer increased mineral deposition

LPL exceeded SPL and 2D controls at days 14 and 21 (Alizarin Red; n=5; p<0.05). Proliferation differences were not significant.

The final bilayer still requires in vivo efficacy validation.

MATERIAL PERFORMANCE

PEU Retained Mass and Mechanical Competence During the Early Healing Window



≈90% mass

remained at day 60 in the reported
in vitro degradation study

6 MPa

dry tensile strength versus 4 MPa
for Cytoflex® ($p=0.0003$)

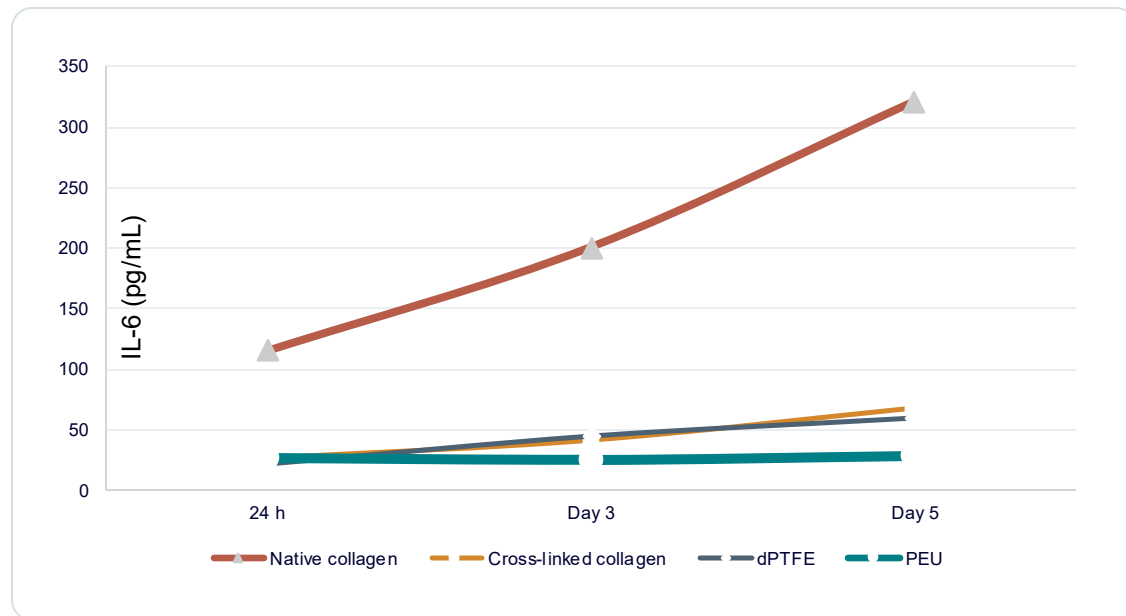
≈120%

elongation at break versus ≈17% for
Cytoflex® ($p<0.0001$)

Interpretation: PEU combined persistence, strength, and conformability—properties relevant to placement and early space maintenance.

COMPLEMENTARY HOST RESPONSE

PEU Elicited a Lower Epithelial IL-6 Response than Native Collagen in-vitro



Stable through Day 5

PEU mean IL-6: 26.9 → 25.1 → 28.3 pg/mL

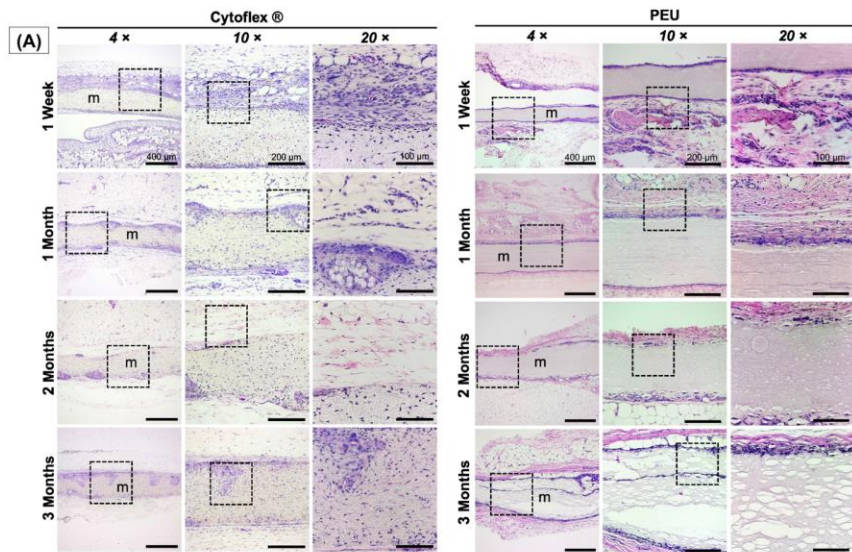
Lower than native collagen

Native collagen increased to 321.0 pg/mL by Day 5; $p < 0.05$ versus PEU at every timepoint.

In vitro epithelial model · n=9 per condition/timepoint

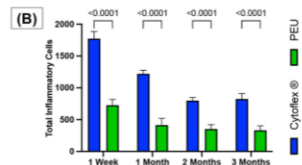
This complementary study supports a low pro-inflammatory epithelial signal; it is not a clinical outcome.

PEU Degradation was Accompanied by a Lower Inflammatory Cell Response



m = membrane

Subcutaneous rat model; single-sheet PEU.

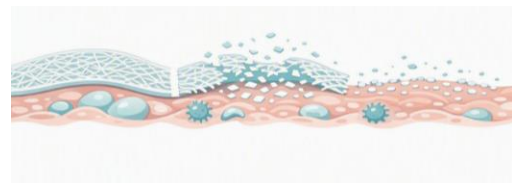


Lower at every timepoint

PEU had fewer inflammatory cells than Cytoflex® from 1 week through 3 months.

$p < 0.0001$

for PEU versus Cytoflex® at each reported interval.



PEU Supported Angiogenesis with Reduced pro-Inflammatory Polarization



CD31 was higher with PEU at 1 month ($p<0.0001$); VWF was higher at 2 months ($p=0.0374$).

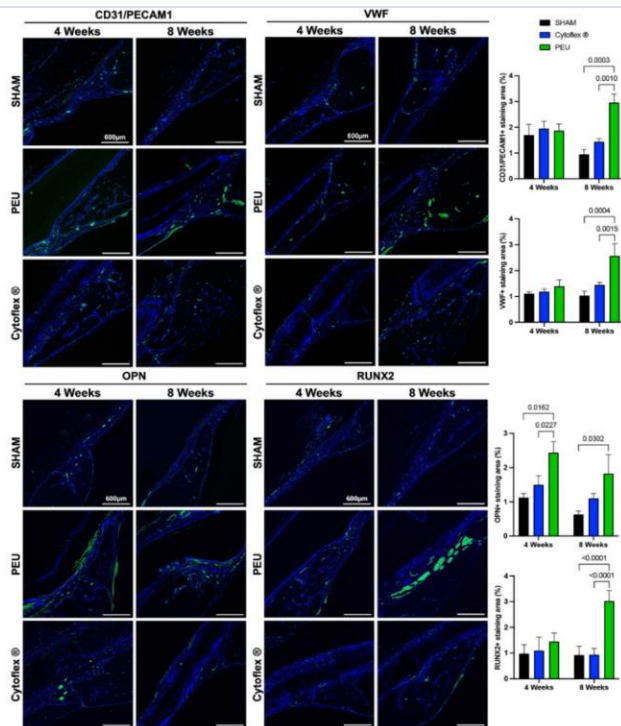
M1 macrophages

iNOS staining was lower with PEU at 2 months (p=0.0003).

M2 macrophages

CD163 differences were not significant at the reported timepoints.

At 8 Weeks, PEU Increased Vascular and Osteogenic Markers within Calvarial Defects



CD31 / PECAM1

Higher than sham and Cytoflex® at 8 weeks
 $p=0.0003$ and $p=0.0010$

VWF

Higher than sham and Cytoflex® at 8 weeks
 $p=0.0004$ and $p=0.0015$

RUNX2

Higher than sham and Cytoflex® at 8 weeks
 $p<0.0001$

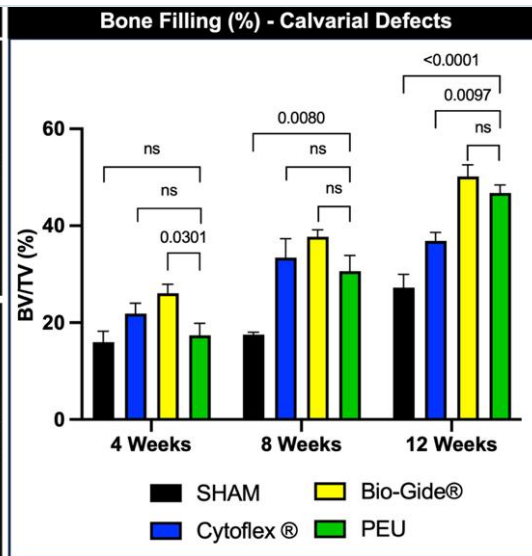
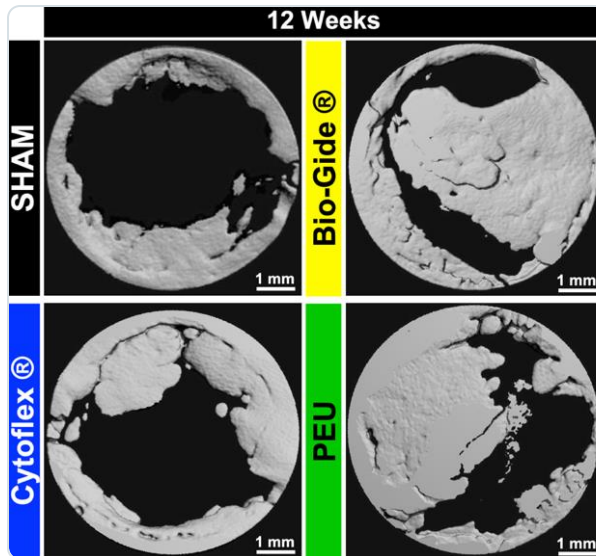
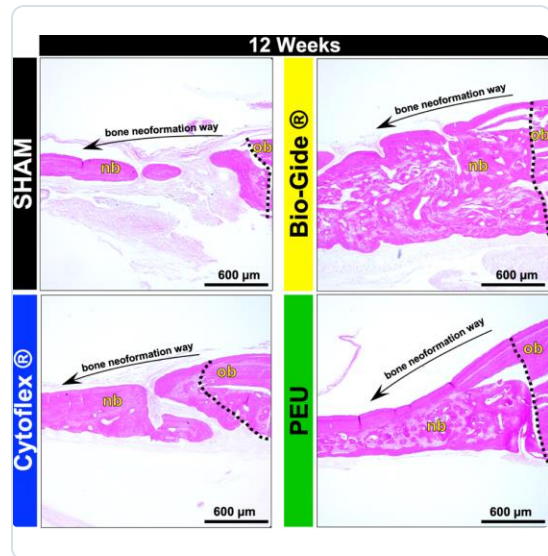
OPN

Higher than sham at 4 and 8 weeks; higher than Cytoflex® at 4 weeks

Mechanistic correlates support—but do not replace—quantitative bone-volume outcomes.

UPDATED COLLAGEN COMPARISON

PEU Performed Comparably to Bio-Gide® at 8–12 Weeks and Exceeded Cytoflex® at 12 Weeks



8 weeks

PEU not different from Bio-Gide® or Cytoflex®

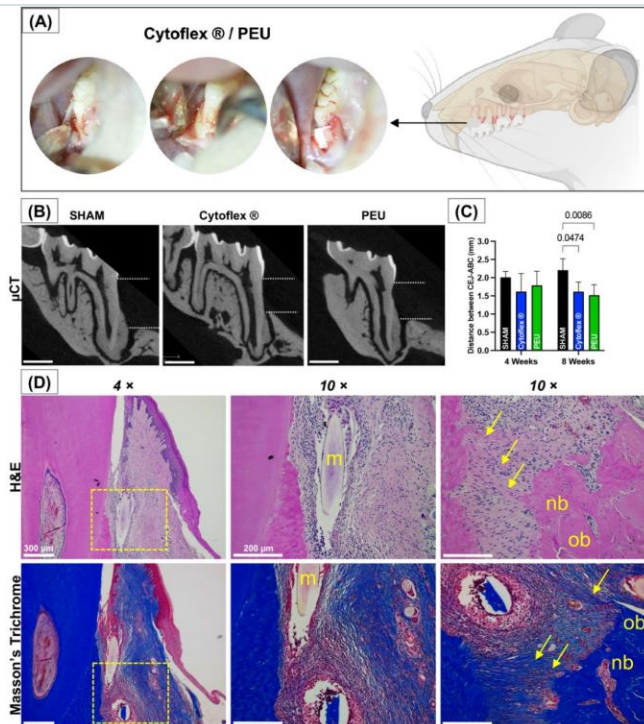
12 weeks

PEU not different from Bio-Gide®
PEU > Cytoflex® ($p=0.0097$)

Bone fill

PEU and Bio-Gide® both > sham at 12 weeks

PEU Restored Alveolar Bone and Supported Periodontal Tissue Regeneration



≈31% recovery

of alveolar bone loss versus sham at 8 weeks;
CEJ-ABC $p=0.0086$.

Functional tissue architecture

Histology showed new cementum and Sharpey's fiber insertion adjacent to newly formed bone.

Barrier function maintained

PEU and Cytoflex® limited soft-tissue ingrowth within the defect.

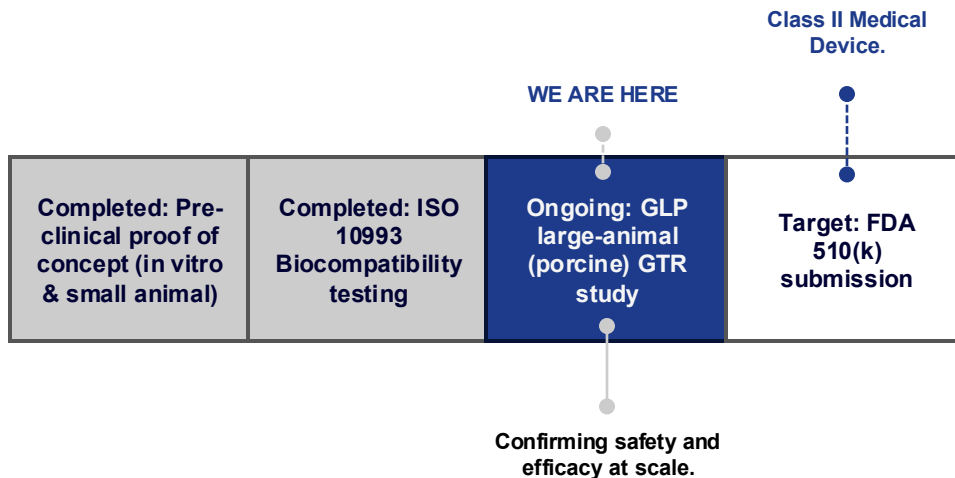
Conclusions and Future Prospectives

Advantages of PEU barrier membrane

Superior mechanical properties	Higher tensile strength, greater elasticity, and improved dimensional stability compared to the commercial membrane.
Excellent cytocompatibility	High cell viability and enhanced osteogenic differentiation of human mesenchymal stem cells.
Lower inflammatory response	Reduced pro-inflammatory cytokine production and favorable immune modulation.
Enhanced angiogenesis	Increased vessel formation and M2 macrophage polarization, supporting tissue regeneration.
Effective bone regeneration <i>in vivo</i>	Significantly more new bone formation and greater bone volume in a clinically relevant calvarial model.

Translational pathway to clinics

Advancing the PEU bilayer membrane through final regulatory milestones to deliver a scalable, clinical-grade solution to the DoW and civilian markets.



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Funding, Collaborators, and Contributors

NIH / NIDCR

Dental barrier membrane program

AFWERX

Broader platform: wound healing, rotator cuff, ophthalmology

U.S. Defense Health Agency

Army collaborator studies

University of Michigan — Prof. Marco C. Bottino group

Renan Dal-Fabbro · Caroline Anselmi · W. Benton Swanson ·
Lais Medeiros Cardoso · Priscila T. A. Toledo · Arwa Daghrery ·
Darnell Kaigler · Alexandra Abel · Marco C. Bottino

Army Postgraduate Dental School / DDEAMC — COL Thomas M. Johnson group

Andrew W. Egger · Denise M. Cacho · Toria L. Koutras ·
Jennifer B. Hawie · Kristen E. Quartararo · Adam R. Lincicum ·
Kimberly A. Inouye · Kenneth M. Hussey · Sanjiv Kumar · Laura
E. Riddle · Michael A. Washington · Michael E. Walsh · Thomas
M. Johnson

With thanks to the Matregenix scientific, engineering, quality, and regulatory teams.

*Army collaborators' views are their own and do not necessarily reflect official policy of the U.S. Government, DHA, or USU.
Key references: Dal-Fabbro et al., ACS Appl. Mater. Interfaces 2024, 16:53419–53434.*

Appendix

Little Innovation in Degradable Polymers in the Last 2 Decades



1971

PGA Dexon™ (Davis & Geck) Absorbable Suture



1987

Polyglyconate Maxon™ (Davis & Geck) Suture



1994

PLA BioScrew™ (Linvatec Corporation) Absorbable Interference Screw
PCL Monocryl™ (Ethicon) Suture



BD Acquired 2021



2005

Polyarylate-Coated TYRX PivitAB™ (TyRx Pharma) Surgical Mesh



2015

Polyurethane Tissueglu™ (Cohera Medical Inc.) Adhesive Glue

1982

PDS™ II (Ethicon) Suture

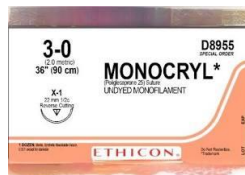


1991

Polyglyconate Suretac™ (Acufex Microsurgical) Absorbable Fixation Device

1996

Polyanhydride Gliadel™ (Gilford Pharmaceuticals) Intracranial Implant



2007

P4HB Tephaflex™ (Tepha Inc.) Surgical Mesh



2020

CBB Citregen™ (Acuitive Technologies) Tendon Interference Screw

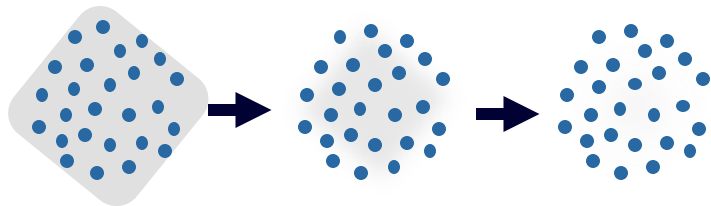
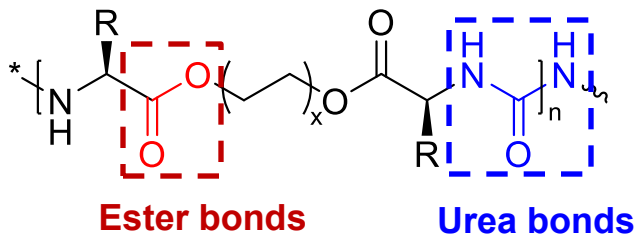


Almost exclusively polyesters
w/ acidic byproducts

Lots of Room &
Demand
For Innovation

Amino-acid-based poly(ester urea): a novel barrier chemistry

Amino Acid-Based Poly(ester urea)s



Uniform degradation & Non-acidic byproducts

Progress in Polymer Science **2024**, 156, 101866.

Advantages

Versatile thermoplastic

- Many validated form factors
- Film, tube, suture, mesh, scaffold

Synthetic Flexibility

- Tunable mechanical properties
- Customizable degradation (weeks to months)

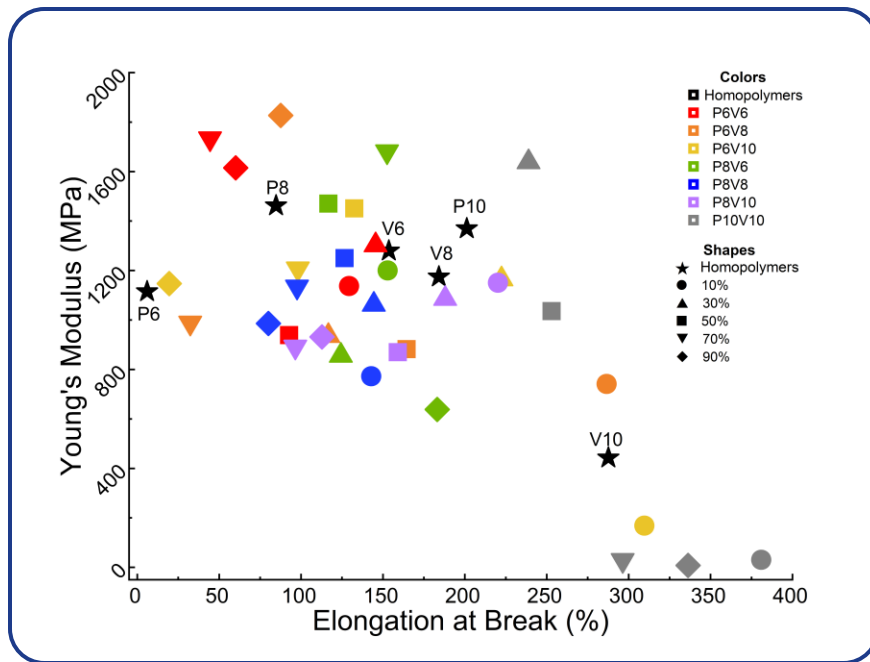
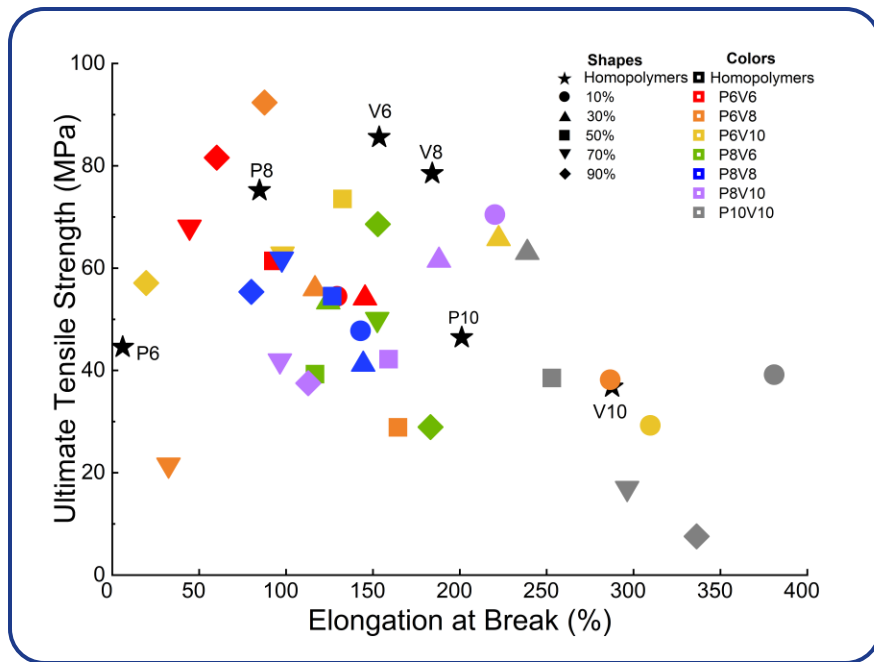
Biocompatible

- Non-toxic byproducts
- Limited inflammatory response

Controlled drug delivery

- Drug stabilization
- Uniform and sustained (days to weeks)

Library of Poly(ester urea)s is Diverse Chemically & Mechanically



CI Dziewior[#], K Godwin[#], NG Judge, NZ Dreger, ML Becker* “Poly(ester urea)s: synthesis, material properties, and biomedical applications” *Progress in Polymer Science* **2024**, 156, 101866.

One Material, Multiple Operational Indications

The amino-acid PEU platform is programmed at the fiber scale — thickness, pore size, and degradation tuned per indication



Dental GBR

Excludes soft tissue;
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Pre-clinical complete



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Full-thickness wound
matrix; ECM-mimetic,
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for load-bearing tendon
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ovine feasibility



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Synthetic "amniotic
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program-stage



Dural repair

Watertight barrier for
neurosurgical defects.

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ONE CHEMISTRY · TUNED ARCHITECTURE · TISSUE FUNCTION

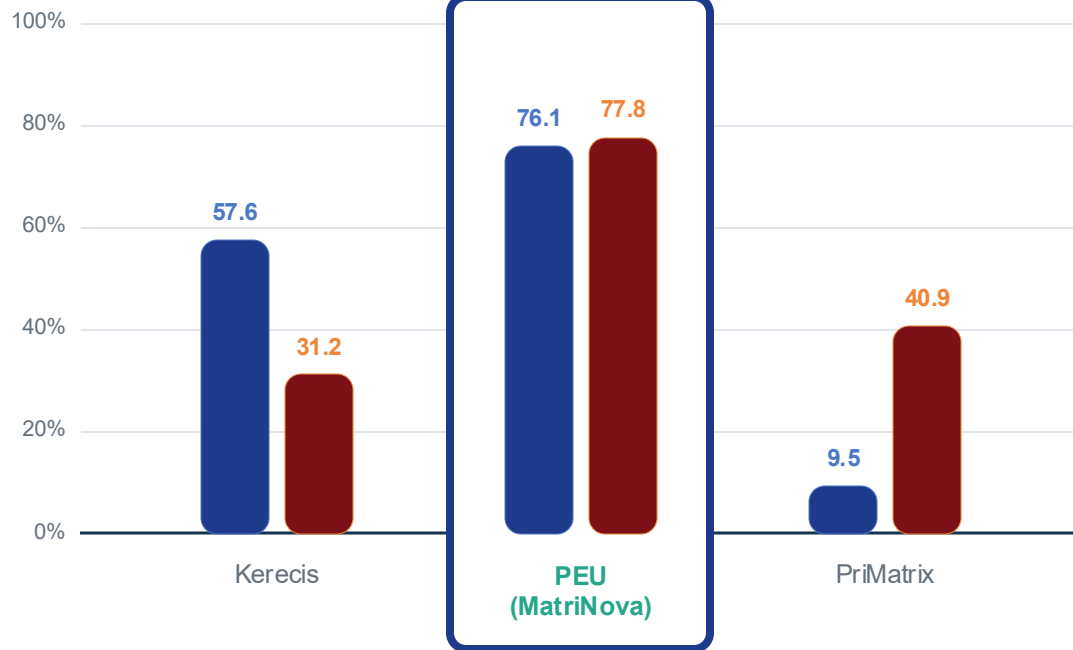
PEU Had the Highest Cell Attachment Against Commercial Skin Substitutes

IN VITRO DERMAL-SCAFFOLD COMPARISON · DESCRIPTIVE SOURCE DATA

Cell attachment to dermal scaffold at Day 7 (%)

Keratinocytes

Fibroblasts



76–78%

attachment on PEU at Day 7

KERATINOCYTES

1.3× / 8.0×

vs Kerecis / vs PriMatrix

FIBROBLASTS

2.5× / 1.9×

vs Kerecis / vs PriMatrix

Wound: PEU-treated Wounds were Smaller at Days 14–28

PIVOTAL PORCINE STUDY · 2 × 2 CM FULL-THICKNESS WOUNDS · n=6 ANIMALS

Representative PEU wound sequence

Separate 4 × 4 cm MatriNova series



Day 0



Day 14

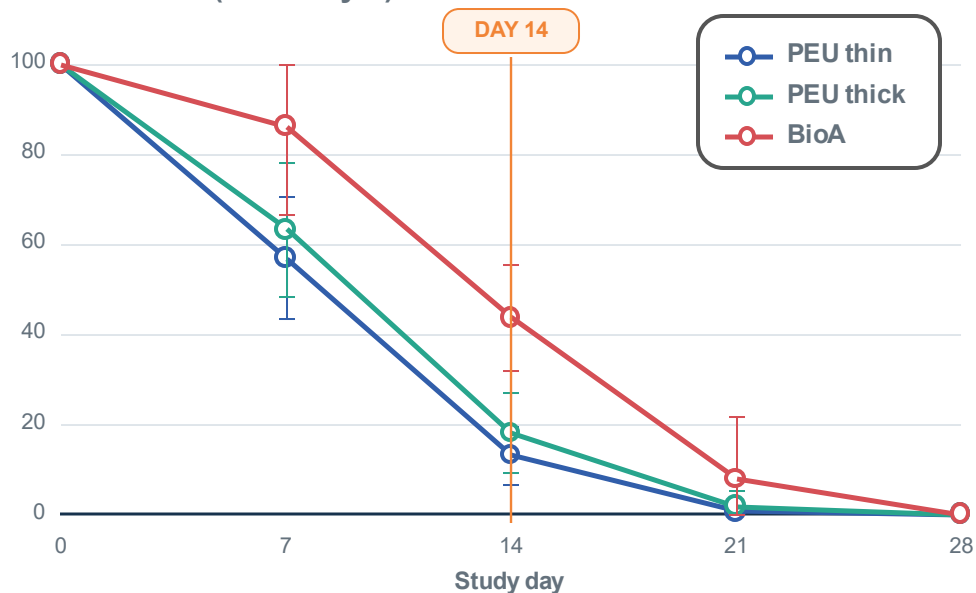


Day 21



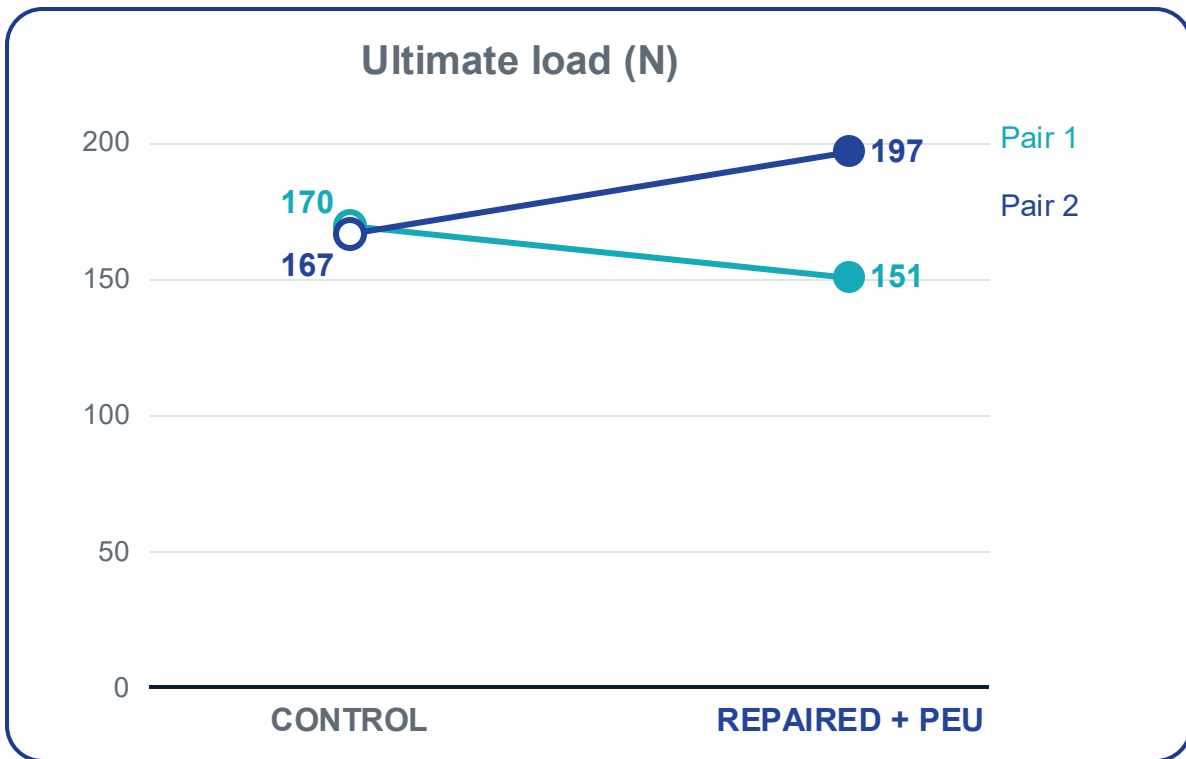
Day 28

Wound area (% of Day 0) · mean $\pm$ SD



DAY 14 · PEU thin 13% · PEU thick 18% · BioA 44% remaining wound area

Rotator Cuff: Repair-site Mechanics Held at 90 Days in Ovine Model



THE REPAIR SITE HELD

Both repaired tendons failed within tendon fibers—not at the surgical repair site.

READ AS FEASIBILITY

**n=2 paired shoulders · 90 days
No statistical power or inferential claim**